EL-NAGERABI, S. A. F., (1) AND ELSHAFIE, A. E. (2)

1-Department of Botany, Faculty of Science, University of Khartoum, PO
 Box 321, Post code 11115, Sudan.

2- Department of Biology, College of Science, Sultan Qaboos University, Sultanate of Oman.

ABSTRACT

In examination of some leguminous crops of the Sudan for the inidence of seed-borne fungi, many species and varieties have been isolated. Nineteen species and 11 varieties are found to belong to the genus Aspergillus, 5 species and 3 varieties (Emericella), 2 species (Eurotium) and one species (Fennellia). From these isolates, 13 species and 8 varieties of Aspergillus together with 4 species and 2 varieties of Emericella are new records to the mycoflora of the Sudan. Some of these species are new records to some of the tested crops. The species are taxonomically described, illustrated and a key to identification is given.

INTRODUCTION

In the Sudan, various seed-crops including different cultivars of legume are cultivated all year round. Under the diversity of vegetation types, climatic conditions and annual rainfall, different saprophytic and pathogenic fungi would be expected to be in association with these crops. Nevertheless, very few to negligible knowledge are known about the mycoflora of the Sudan. Very little studies were conducted on rusts, smuts, powdery mildews and leaf spotting diseases (Tarr, 1955, 1963). Agaricus, polypores, coprophilous fungi, coelomycetes and many of dematiaceous hyphomycetes have not yet investigated, and no attempt has been focused to describe them taxonomically. Only some species of *Curvularia* and *Drechslera* have been studied (Elshafie, 1985, 1986). This study is a series of papers in which some of seed-borne dematiaceous hyphomycetes will be

described. In this paper, Aspergillus species and their teleomorphs isolated from the seeds of Sudanese leguminous crops are described, illustrated and a key for identification is given.

MATERIALS AND METHODS

Collection of Seed Samples

In the present test, 104 seed samples from 8 of the Sudanese legumes viz: Guar (Cyamopsis tetragonoloba), Soybean (Glycine max), lentil (Lens esculenta), lupine (Lupinus termis), pea (Pisum sativum), fenugreek (Trigonella foenum-graecum), faba bean (Vicia faba) and cowpea (Vigna unguiculata) were purchased from the local markets of Khartoum and supplied by Arab Organization for Agricultural Development (AOAD) from Agadi Farm (Damazin, Blue Nile State) and Um Dom Farm (Khartoum State), National Centre for Research and from Guar Production Company. The samples represent the harvesting seasons of 1993-1996. The working samples were drawn and examined according to the International Rules for Seed Testing Association (ISTA, 1966).

Isolation of seed-borne Aspergilli

For the isolation of seed-borne Aspergilli, routine Agar Plate and Blotters (Moistened Chambers) Methods were adopted (Christensen, 1963; Hussain et al., 1989; Zohri and Abdel Gawad, 1992; El-Kady and Youssef, 1993; Moslem and Parvez, 1993). In these method, 800 seeds from each samples were disinfected with mercuric chloride (0.1%, 5 minutes) and inoculated aseptically on Potato Dextrose Agar (PDA) and moistened filter papers (Blotter). The inoculated plates were incubated at 28 ± 2 °C for 1-2 weeks and were then examined using stereoscopic binocular microscope to determine the natural growth of fungi on the seeds.

Identification:

The identification of the genus Aspergillus and their teleomorphs was carried out with the aid of the microscope, whenever possible on the seeds. When this was not possible, the isolated fungi were inoculated on various diagnostic growth media. The isolates were incubated at 25 and 37 °C on Czapek Dox Agar (CDA) (Singh et al., 1991; Moubasher, 1993), and at 25 °C on Czapek Yeast Extract Agar

(CYA) (Pitt, 1973), Czapek Yeast Extract Agar with 20% Sucrose (CY20S) (Pitt and Hocking, 1985) and Malt Extract Agar (MEA) (Blakeslee, 1915).

The identification of the isolated species was confirmed using many taxonomic papers and monographs (Raper and Fennell, 1965; Raper and Thom, 1968; Klich and Pitt, 1988; Singh et al., 1991; Moubasher, 1993). Different publications of Commonwealth Mycological Institute (CMI), International Mycological Institute (IMI) and Danish Government Institute of Seed Pathology for Developing Countries were also used.

RESULTS AND DISCUSSION

Microscopic examination of the different seed samples of the leguminous crops inoculated on Potato Dextrose Agar (PDA) and moistened filter papers (Moistened Chambers) at $28 \pm 2^{\circ}$ C revealed that the seeds of these crops were infected with 19 species and 11 varieties of the genus Aspergillus and 3 species and 3 varieties (Emericella), 2 species (Eurotium) and one species (Fennellia). From these isolates 12 species and 8 varieties of Aspergillus, and 4 species and 2 varieties of Emericella and one species (Fennellia) are new reports to the mycoflora of the Sudan. Some of these isolates are new records to some of the tested crops. Occurrence of this large number of fugal species and varieties under the warm conditions of the Sudan proved their adaptation to this climate as suggested by Elshafie (19985, 1986).

It is evident that the seeds of leguminous crops are infested with numerous fungi some of which are new to these crops and to the mycoflora of the Sudan. The study revealed that the seeds of these crops harbour many of the devastating pathogens which proved destructive under favourable field and storage conditions. This indicates that under the warm conditions of the Sudan different crops would be expected to be infected with many new fungal species and even new genera and taxonomic groups. Therefore the seeds for sowing purpose should be certified, treated and stored under proper conditions to reduce the spreading of these pathogens to uninfected areas. Hence, the construction of proper quarantine stations in the Sudan is badly needed. Various factors are known to cause a considerable fluctuation in the seed-borne mycoflora, therefore timely

| Some of these fungi are known to produce secondary metabolite the seeds which are found to be toxic to both man and his dome livestock. Therefore, continuous assessment is needed to prevent consumption of such contaminated seeds. Key to the seed-borne species of Aspergillus, Emericella Eurotium is based on that of Klich and Pitt (1988) and adopted | stic the |
|--|-------------|
| Moubasher (1993). | 2 |
| 1- Aspergilla predominately biseriate | 24 |
| 1- Aspergilla predominantly uniseriate | 24 |
| BISERIATE SPECIES:- | |
| 2- Metulae present or absent, with the former case predominant; conidial heads yellow green, radiate to very loo columnar | us only |
| 3- Conidial heads in compact column | 4 |
| Conidial heads radiate to loosely columnar | 5 |
| 4- Sclerotia-like masses produced on Malt Extract A (MEA) | reus ves |
| 5- Conidia on CYA greyish-turquoise to dark turquoise | A. |
| sydowii | 7 |
| 6- Conidia smooth to finely roughened 6- Conidia consistently roughened when mature | 8 |
| 7- Colonies on MEA greyish-yellow, sporulation often sparse, | |
| largest conidia often up to 7 µm diameter | yzae |
| 7- Colonies on MEA grey green, largest conidia up to 3.5 µm in | |
| diameter | llifer |
| 8- Colonies dark brown to black on CYA | 9 |
| 8. Colonies dull green to grev green CYA | 10 |
| O Largest conidia < 6 um in diameter | uger |
| 9- Largest conidia > 6 µm in diameter | uus |
| 10- Stipes brown, maximum length 200 μm in length | A. |
| tetrazonus 99 | |

| 10- Stipes sinnamon brown, maximum length 200 μm in length |
|--|
| 11- Colonies on MEA dark brown to black, largest |
| conidia 5 µm in diameter |
| 11- Colonies on MEA golden yellow dark green |
| largest conidia 5 μm in diameter 13 |
| 12- Conidiophores > 700 μm in length |
| 13- Conidiophores maximum length 500 μmA. terreus var aureus |
| 13- Conidiophores maximum length < 200 µm |
| 14- White, spicular hyphae arising above the conidial heads, readily visible under stereoscopic microscope |
| 14- White spicular hyphae absent |
| 15- Cleistothecia absent |
| 15-Cleistothecia present Fennellia flavinos |
| 16- Stipes finely roughened to very rough, colorless, |
| maximum length 500 μm |
| 16- Stipes thick, usually, smooth-walled, brown in age, |
| maximum length 500 μm |
| 17- Cleistothecial envelope in dull shades consisting of hulle |
| cell only |
| 17- Cleistothecial wall consisting of interwoven hyphae, |
| but has an envelope of hulle cellsA. nidulans var. echinulata |
| 8- Slow growth on all media |
| 8- Moderate to rapid growth |
| 19- Ascospores with 4 short longitudinal flanges not |
| always readily visible |
| 9- Ascospores with less than 4 longitudindal flanges |
| always readily visible 20 |
| 20- Ascospores convex wall echinulate; orange red in |
| colorE. nidulans var. echinulata |
| 0- Ascospores convex wall echinulate, lenticular, |
| violet blue in color |
| 1-Ascospores stellateE. variecolor |
| 1-Ascospores not stellate 22 |

| 1. 11llow hyphae hearing | |
|--|--|
| 22- Cleistothecia surrounded by yellow hyphae bearing | nidulans |
| hülle cells, dark red at maturityE. nidulans var. | manans |
| 22- Cleistothecial wall composed of hülle cells only, | Lata |
| colored | var. iaia |
| UNISERIATE SPECIES:- | |
| 23- Osmophilic, colony diameter on CY20S more | |
| than twice on CYA | 24 |
| 23- Not osmophilic, colony diameter on CY20S less | |
| than twice on CYA | 25 |
| 24- growth on CYA at 37°C very low, diameter | |
| < 15 mm | llandicus |
| < 15 mm | hrunneus |
| 24- No growth on CYA at 3/°C | 26 |
| 25- Conidial heads columnar | 27 |
| 25- Conidial heads radiate splitting into columns with age | .21 |
| 26. Vesicles mostly pyriform or spathulate; conidia | |
| nradominantly globose echinulate greenish | Compaire ortage |
| to green in mass | <i>fumigaius</i> |
| 26 Vasiala subalabase conidia globose to subglobose | |
| smooth to echinulate, yellow green | olumnaris |
| 27 Conidia predominantly ellipsoidal, maximum | |
| vesicles diameter $> 45 \mu \text{m}A$, japonicus var. α | aculeatus |
| 27 Canidia globose to subglobose, maximum | |
| vesicles diameter < 45 µm | japonicus |
| 28- Yellow cleistothecia present (Eurotium) | 29 |
| 28- Yellow cleistothecia absent | 30 |
| 29- Convex surface of ascospores roughened, | |
| with conspicuous ridge of irregular height | |
| on either side | ıstelodami |
| oo Carrier surface of ascospores smooth, low | |
| or inconspicuous longitudinal ridges | E. rubrum |
| 30- Conidia hyaline when yong and brown with | |
| age, evenly space spines | . japonicus |
| 30- Conidia yellow green, smooth echinulate | A. flavus |
| 30- Comma yellow green, smooth commander | , and the second |
| var. columnaris The descriptions of the isolates are based on cultures in | ncubated at |
| The descriptions of the isolates are based on cultures in | Moubasher |
| 25 and 37°C on Czapek Dox Agar (Singh et al., 1991, I | % Sucrose |
| 1993) and at 25 °C on Czapek Yeast Extract Agar with 20 | ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,, |

(Pitt and Hocking, 1985) and Malt Extract Agar (Klich and Pitt, 1988). The inoculated plates were incubated for 7 days in the dark (Pitt, 1973).

A. caespitosus Raper and Thom (Fig. 1).

On CYA colony 30-45 mm in diameter; at 37 °C 10-29 mm, low, velvety, sulcate; conidial area greyish-green; mycelium white to pale yellow; soluble pigment absent; reverse brown exudate on the surface; reverse wrinkled or rhizoid, red brown. On CY20S 30-53 mm in diameter; the appearance similar to that. On CYA on MEA 25-40 mm in diameter low, dark green; mycelium inconspicuous, white or yellow brown near the hülle cells; reverse yellow to red brown; margin fimbriate. On CDA 33-47 mm in diameter, appearance as on CYA. Heads radiate on CYA and loosely columnar on MEA. Stipes smooth, brown with age, 45-150(commonly 61.2 m) in length. Vesicles spathulate to hemispherical, 7.5-15.6 m(commonly 12.5-15 mm) wide. Metulae $4.1-10 \times 2.5-3.5$ m (commonly 3.0m)wide. Phialides ampulliform, $5.0-8.8 \times 2.5-4.1 \mu m$ (commonly 3.0-4.1 mm) Conidia spherical, rough-walled, greyish-green 3.0-5.2 mm in diameter(commonly 5mm). Ascomata absent; hülle cells hyaline to reddish with age.

The fungus has been isolated from the seeds of soybean ,pea , fenugreek and faba bean .

A. flavus var. flavus Link. (Fig. 2).

On CYA colony diameter 40-67 mm, floccose, olive green to olive yellow; sclerotia dark brown to black; exudate present; reverse colorless to dark brown. On CY20S 54-73 mm in diameter, other characters similar to those on CYA except exudate absent and reverse bright yellow. On MEA 53-65 mm in diameter, floccose, olive to dark green; mycelium white, thin; exudate absent; reverse colorless. On CDA 45-55 mm in diameter, floccose, yellow green; margin white, thin; reverse colorless. Heads radiate to columnar

^{8.}

[:] New records for the crop (s).

with age. Stipes hyaline, verrucose, flexuous or straight, 533-635.1 x 8.2-12.3 μ m. Vesicles globose to subglobose or flask-shaped, fertile on the entire surface, hyaline to yellowish, 25.6-46.2 μ m in diameter. Metulae absent in small heads, present or absent in large ones, 5.5-143 \times 5.0-8.4 μ m. Phialides ampulliform, 7.4-10.3 \times 3.3-4.1 μ m. Conidia globose to subglobose echinulate, yellowish-green, 3.2-5.1 $\acute{y}\mu$ m in diameter. Ascomata absent.

The fungus has been isolated from guar, soybean, lentil, lupine, pea, fenugreek, faba bean and cowpea.

A. fumigatus Fresenius (Fig. 3)

On CYA colony diameter 34-58 mm, at 37° C 57-66 mm, velutinous to floccose, plane or furrowed radially, greyish-turquoise to dark turquoise, mycelium white, exudate colorless; reverse yellow to red brown; soluble pigment absent. On CY20S 33-62 mm in diameter; colors and other characters as on CYA. On MEA 37-65 mm in diameter; conidial heads colors as on CYA, mycelium white, inconspicuous; reverse colorless to pale yellow; soluble pigment colored as reverse. On CDA 50-54 mm in diameter; velutinous, bluish-green to dark grey with age; reverse colorless to creamish-yellow. Heads mainly columnar, compact. Stipes smooth, colorless to greenish-grey, $350-500 \times 5.3-8.2$ mm. Vesicles spathulate, 12.3-28.6 µm wide. Metulae absent. Phialides ampulliform, closely packed, roughly paralleled to the axis of the stipe $7.3-8.0 \times 2.13.0$ µm. Conidia globose, echinulate, greenish to green in mass 3.3-4.1 µm. Ascomata absent.

The fungus has been recovered from the seeds of guar, soybean, lentil, lupine, pea, fenugreek, faba bean and cowpea.

A. hollandicus Samson and W. Gams. (Eurotium amstelodami Mangin). (Fig. 4).

On CYA colony diameter 8-22 mm, at 37°C 9-13 mm, low plane, grey green to greyish-turquoise; mycelium white to yellow; reverse yellow to brown; soluble pigment brown. On CY20S 39-52 mm in diameter, dark green; mycelium colored as on CYA; reverse yellow beneath cleistothecia or green beneath conidia. On MEA 10-19 mm in diameter; dark green; mycelium inconspicuous; cleistothecia absent; reverse colorless. On CDA 15-25 mm in diameter; floccose, green

103

yellow, exudate absent; margin thin, yellow; reverse brown to tan brown. Heads radiate to loosely columnar. Stipes colorless to pale yellow, $280\text{-}330 \times 9\text{-}12.5~\mu\text{m}$. Vesicles alobose, fertile on the upper half, $10\text{-}27~\mu\text{m}$ wide. Metulae absent. Phialides ampulliform with broad neck, $6.3\text{-}8.5 \times 2.5\text{-}5.0~\mu\text{m}$. Conidia globose to subglobose or barrel-shaped with flattened ends, finely spinulose, $2.5\text{-}6.5~\mu\text{m}$ in long axis. Ascomata cleistothecia globose to subglobose, yellow, $115\text{-}120~\mu\text{m}$ in diameter; asci $10.0\text{-}12.0~\mu\text{m}$; ascospores lenticular, V-shaped equatorial furrow, two irregular ridges, rough convex wall, $3.8\text{-}5.0 \times 3.5\text{-}4.3~\mu\text{m}$.

The fungus has been reported on the seeds of lentil and pea.

A. japonicus Saito var. aculeatus (Lizuka) Al-Musallam. (Fig. 5).

On CYA colony diameter, 55-77 mm, at 37°C 13-25 mm; conidial area red brown to black with age; mycelia white with central floccose overlay; reverse peach yellow to brown; exudate absent; sclerotia absent. On CY20S 68-83 mm in diameter; other characters similar to those on CYA. On MEA 40-67 mm in diameter; conidial area dark brown to black, sparse sporulation; reverse yellowish. On CDA 45-67 mm in diameter; conidial area black; mycelium white; reverse colorless. Heads radiate, splitting into divergent columns with age. Stipes smooth, colorless or brown near the apices, 250-500 × 10-15 μ m. Vesicles elongate to globose, collapsing near the base, 25-85 μ m wide. Metulae absent. Phialides ampulliform with flat base, covering the three quarters or the whole vesicle, 5.0-12.0 × 2.5-5.0 μ m Conidia predominant ellipsoidal, globose, widely spaced spines, 4.5-5.5 × 3.5-4.5 μ m. Ascomata absent.

The fungus has been reported on the seeds of lupine.

A. japonicus Saito var. japonicus (Fig. 6).

On CYA colony diameter 45-52 mm, at 37 °C 33-42 mm; conidial area very dark brown; mycelium white; exudate and soluble pigment absent; reverse dull brown. On CY20S 62-74 mm in diameter; other characters similar to those on CYA. On MEA 55-64 mm in diameter; conidial area dark brown. On CDA 40-54 mm in diameter; conidial area black; exudate absent; reverse colorless. Heads radiate or spilt into few loose columns. Stipes smooth, hyaline below, slightly pigmented at the apices, sinuous, 150-350 × 8.2-10.0 µm. Vesicles globose, pigmented, fertile over the entire surface in

small heads, but at least the half of the surface, 10-43.0 um. wide. **Metulae** absent. **Phialides** ampulliform, $6.0\text{-}8.0 \times 3.5\text{-}4.0 \mu\text{m.}$ **Conidia** globose to subglobose, hyaline when young and brown with age, evenly spaced spines, $3.0\text{-}5.0 \mu\text{m.}$ **Ascomata** absent.

The fungus has been isolated from the seeds of lupine and fenugreek.

A. nidulans (Eidam) Winter var. echinulata (Emericella nidulans (Eidam) Vuillemin var. Echinulata (Raper and Fennell) Subramanian) (Fig. 7).

On CYA colony diameter 42-50 mm, at 37°C 44-58 mm, velutinous, plane to slightly sulcate, green to deep green; mycelium white to cream; exudate red to brown; reverse brown orange to purple red. On CY20S 33-45 mm in diameter; other characters similar to those on CYA. On MEA 40-55 mm in diameter, colony low, plane, heavily sporulated, dark green; mycelium white, inconspicuous; reverse pale brown. On CDA 17-32 mm in diameter; conidial area dark grey to dark brown raised, granulated; exudate faint droplets; reverse brown, tan brown to reddish-brown. Stipes short, smooth, golden brown, 61.2-175.4 mm in length. Vesicles globose to subglobose, fertile on the upper half, 12.2-16.4 µm in diameter. Metulae each one bearing 1-2 phialides, $4.0-6.5 \times 2.5-3.1$ µm. Phialides ampulliform with very short neck, $5.0-8.2 \times 2.5-4.1$ subglobose, thick, finely echinulate, μm. Conidia globose to greenish, 2.5-4.4 µm in diameter. Ascomata cleistothecia reddishblack to black, 110-320 µm; asci ovate to subglobose, orange red; ascospores lenticular, two equatorial ridges, convex wall echinulate, 2.5-6.3 um in diameter.

The fungus has been isolated from the seeds of pea and fenugreek.

A. nidulellus Samson and W. Gams var. nidulans (E. nidulans). Vuillemin var. nidulans) Eidam (Fig. 8).

On CYA colony diameter 40-55 mm, at 37°C 47-66 mm, velutinous or with floccose overlay, plane or sulcate, green to deep green; mycelium white to cream; exudate dull pale red to brown; reverse brown orange to deep purple red; soluble pigment similarly colored. On CY20S 38-48 mm indiameter; conidial and mycelial colors similar to those on CYA; exudate absent; reverse red brown;

105

soluble pigment absent. On MEA 45-62 mm in diameter; colony low, velutinous, plane, heavily sporulating with dark green conidia; mycelium white, inconspicuous; cleistothecia dull yellow to buff; reverse pale brown. On CDA 36-43 mm in diameter; conidial area yellow green mixed with yellowish to black cleistothecia, exudate absent; reverse tan brown to reddish-black brown. Heads radiate to loosely columnar. Stipes smooth, brown with age, sinuous, 83.8-98.4 \times 8.0-12.5 µm. Vesicles spathulate to hemispherical, fertile on the upper half, 4.2-12.3 µm. Metulae bearing two phialides each, 6.6-7.0 \times 2.5-3.1 µm. Phialides ampulliform with very short neck, 5.0-6.3 \times 2.5-3.3 µm. Conidia globose, greenish to deep green, echinulate, 2.5-4.1 µm in diameter Ascomata cleistothecia dark red with age, 143.5-266.5 µm in diameter; hülle cells 12.3-24.6 µm in diameter; ascospores lenticular, 2equatorial crest smooth convex walls purplish-red, 5.0 - 7.5 \times 2.0-3.8 m.

The fungus has been recovered from the seeds of guar, soybean, lentil, lupine, pea, fenugreek, faba bean, and cowpea.

A. niger Van Tieghem (Fig. 9).

On CYA colony diameter 50-73 mm, at 37°C 55-68 mm, floccose, sulcate due to the varying length of stipes; conidial area very dark to black; mycelium white to dull yellow, inconspicuous; reverse vellow. On CY20S 56-64 mm in diameter, colony appearance similar to that on CYA with more densely packed aspergilla. On MEA 39-58 mm in diameter; conidial area black; colony granular to floccose; mycelium white and inconspicuous; reverse colorless.On CDA 55-68 mm in diameter; conidial area blackish-brown to carbon black to naked eye; mycelium yellow; margin white to pale yellow; reverse yellow to creamish yellow. Heads globose splitting into many irregular columns with age. Stipes smooth, thick, hyaline, yellowish to brownish near the apices, 450- $500 \times 13\text{-}15 \ \mu\text{m.}$ Vesicles spherical to subglobose, hyaline to brownish, 50-76.5 µm wide. Metulae long, closely packed, brownish, $11\text{-}13.8~\times~4.2\text{-}5.5~\mu m.$ Phialides short, thick, ampulliform, $5.0\text{-}6.2~\times~4.2\text{-}5.5~\mu m.$ 2.5-3.2 µm. Conidia globose to elliptical, verruculose, with irregular ridges and bars, dark brown to black, 4.0-5.0 µm in diameter. Ascomata absent.

The fungus has been isolated from the seeds of guar, soybean, lentil, lupine, pea, fenugreek, faba bean and cowpea.

A. niger Van Tieghem var. awamori (Nakazawa) AL-Mussalam (Fig. 10).

On CYA colony diameter 58-75 mm, at 37°C 52-67 mm, velutinous, low, brown to black; mycelium white; exudate absent; reverse dull yellow to dull brown. On CY20S 65-77 mm in diameter; other characters similar to those on CYA. On MEA 48-73 mm in diameter; dark brown to black, plane; mycelium inconspicuous; aspergilla uncrowded. On CDA 60-74 mm in diameter; conidial area blackish-brown; mycelium white; reverse pale yellow to colorless. Heads radiate, splitting into columns at maturity. Stipes smooth, thick, colorless to pigmented near the apices, $350-750 \times 8.0-14.0 \mu m$. Vesicles globose to subglobose, fertile over the entire surface or upper half in small vesicles, $7.5-45.0 \mu m$. Metulae hyaline to subhyaline, $5.0-9.0 \times 4.0-7.50 \mu m$. Phialides hyaline to light brown; $5.0-10.0 \times 3.5-4.0 \mu m$ Conidia globose to subglobose, hyaline to light brown, smooth to delicately roughened, ornamented with ridges and bars, $4.5-6.0 \mu m$ in diameter. Ascomata absent.

The fungus has been reported on soybean, lentil, lupine, fenugreek and cowpea.

A. oryzae (Ahlburg) Cohn (Fig. 11).

On CYA colony diameter 60-75 mm, at 37°C 45-65 mm, colonies lanose to floccose, greyish-yellow to olive brown, mycelium dense, white to cream, exudate absent; reverse colorless. On CY20S 65-73 mm in diameter; colonies floccose, olive yellow to greyish yellow; mycelium white, dense; conidia abundant; reverse colorless. On MEA 55-60 mm in diameter; colonies floccose, greyish-yellow; mycelium white and not dense as on CYA; conidia sparse; reverse colorless. On CDA 49-63 mm in diameter, colonies floccose, yellowish-green in the margin to grey green in the centre; exudate absent; reverse colorless. Heads radiate to loosely columnar. Stipes rough-walled, colorless, $200-1000 \times 8.0-12.0 \mu m$. Vesicles clavate to subglobose, 20-48 µm wide. Metulae commonly absent (uniseriate) or biseriate with both conditions present on the same head, $5.0-8.0 \times$ 3.0-4.5 µm. Phialides covering the most of the vesicle surface, 8.0- 15.0×3.0 -5.0 μm . Conidia smooth to finely roughened, globose, ovoid, ellipsoidal, 3.0-7.0 µm in diameter. Ascomata absent.

The fungus has been isolated from the seeds of guar, soybean, lentil, lupine, pea, fenugreek, faba bean and cowpea.

A. rubrobrunneus Samson and W. Gams. (E. rubrum Konig, Spieckermann and Bremer) (Fig. 12).

On CYA colony diameter 25-33 mm, at 37°C no growth, velvety, plane with umbonate centre, conidial heads sparse, grey green; mycelium and cleisothecia forming yellow felt; exudate absent; reverse brown. On CY20S 43-67 mm in diameter, plane or radially sulcate, velutinous with floccose overlay, cleistothecia and mycelium yellow; reverse bright. On MEA 17-55 mm in diameter, granular or tuft, sparse, grey green; mycelium yellow gold; cleistothecia rare, yellow gold; reverse yellow. On CDA 30-42 mm in diameter, velutinous, plane, greyish-green; mycelium and cleistothecia pale reverse brown to dark brown. Heads radiate on CY20S. vellow felt; Stipes smooth to rough, colorless to pale brown, 100-500 ×3.0-16.0 μm. Vesicles subglobose, 7.0-32.8 ýμm. Metulae absent. Phialides ampullifrom, covering most of the vesicle, $7.5-14.0 \times 2.5-5.0 \,\mu m$. Conidia variable in shape, spherical, ellipsoidal, apiculate, spinose, $4.5-6.5 \times 4.0-6.0 \ \mu m$. Ascomata cleistothecia, subglobose, wall of single layer of pseudoparenchyma, asci 9.5-17.0 µm, ascospores lenticular, hyaline, slight furrow, inconspicuous ridges, smooth surface, $5.0\text{-}6.0 \times 3.0\text{-}4.5 \ \mu m$.

The fungus has been reported on the seeds of lentil.

A. stellifer Samson and W. Gams (E. variecolor Berkeley and Broome (Fig. 13).

On CYA colony diameter 30-45 mm, at 37°C 15-25 mm, velutinous, grey green; mycelium submerged, sparce; margin white; exudate absent; cleistothecia yellowish grey; reverse yellow green to black at the centre. On CY20S 25-35 mm in diameter, others characters similar to those on CYA except that the colony umbonate, green to yellowish-green. On MEA 40-55 mm in diameter, plane, zonate; conidial area and cleistothecia as on CYA; exudate absent; reverse dull yellow. On CDA 15-25 mm in diameter, velutinous, green to grey green; margin white, cleistothecia yellowish; reverse yellow green. Heads radiate to loosely columnar. Stipes smooth, cinnamon brown, $150-260 \times 3.5-5.5 \mu m$. Vesicles hemispherical, 8.2-12.3 µm. Metulae each one bearing 2-3 phialides,

6.5-8.4 \times 3.2-4.4 μm. **Phialides** ampulliform, 7.4-8.4 \times 2.1-3.3 μm. **Conidia** globose, rugulose, greenish to green, 2.5-3.5 μm. **Ascomata cleistothecia** two type; aggregated 350-550 μm, segregated 150-250 μm; **asci** subglobose or elongated, stellate, 10-12 \times 8.0-10 μm; **ascospores** lenticular, bearing two equatorial ridges, 2.0-3.5 μm wide, stellate, ascospore body 3.0-4.0 μm face view, 3.0-4.5 \times 2.0-3.0 μm side; **hülle cells** globose to irregular, 15.4-25.0 μm in diameter.

The fungus has been reported on the seeds of fenugreek

A. sydowii (Bain. and Sart.) Thom and Church (Fig. 14).

On CYA colony 20-35 mm in diameter, at 37°C 8-12 mm, velutinous, sulcate or radially wrinkled, umbonate, dark turquoise to dark green; margin white; exudate dark brown to reddish-brown; reverse red brown to reddish-black. On CY20S 22-35 mm in diameter, characters similar to that on CYA except exudate absent and texture somewhat floccose. On MEA 15-20 mm in diameter, color as on CYA, granular to floccose, plane; exudate absent; mycelium white; margin white; reverse colorless. On CDA 13-18 mm in diameter, conidial area blue green to green, raised or umbonate, radially wrinkled; exudate present; reverse grey orange with light pigment diffusing into the media. Heads typical radiate to globose. Stipes thick-walled, colorless, $100-380 \times 2.0-8.2 \mu m$. Vesicles globose toclavate, fertile on the entire surface, 2.5-20.0 µm in diameter. Metulae broad, pyriform, divergent, 3.0-5.2 × 2.0-3.5 μm . Phialides ampulliform, with broad neck, 4.2-7.4 \times 2.1-3.2 μm . Conidia globose to subglobose, echinulate, to spinose, 3.5-4.2 µm in diameter. Ascomata hülle cells produced in a few numbers.

The fungus has been isolated from the seeds of guar, soybean, lentil, pea, faba bean and cowpea.

A. terreus Thom var. terreus (Fig. 15).

On CYA colony 28-46 mm in diameter at 37°C 37-86 mm velutinous to lanose, plane to radially sulcate, brownish-orange; mycelium white; pigment absent; reverse yellow to pale brown. On CY20S 30-69 mm in diameter, color similar to that on CYA, sulcate, floccose. On MEA 37-57 mm in diameter; conidia sparce, pale orange; mycelium white, inconspicuous; reverse yellow to pale brown. On CDA 12-32 mm in diameter, velutinous, sulcate, umbonate, cinnamon or golden yellow; reverse yellowish-brown with

yellowish secretion in the media. Heads compact columnar. Stipes smooth, hyaline, straight or flexuous, 80-300 µm long. Vesicles spherical to hemispherical, fertile on the upper two third, 10.4-16.4 μm in diameter. Metulae almost parallel to the stipes, cylindrical, 5.0- 6.5×2.0 -2.5 µm. Phialides thin, 5.4- 6.9×1.3 -2.3 µm. Conidia globose to subglobose, in long chains, $1.5-2.7~\mu m$ in diameter. Ascomata absent.

The fungus has been recovered from the seeds of guar, soybean, lentil, lupine, pea, fenugreek, faba bean and cowpea.

A. tetrazonus (Thom and Raper) Samson and W. Gams (E. quadrilineata (Thom and Raper) Benjamin (Fig. 16).

On CYA colony 45-50 mm in diameter, at 37°C 50-65 mm, velutinous, low, radially sulcate, dull green to grey green; mycelium white; exudate absent; reverse red brown. On CY20S 55-62 mm in diameter, low, sulcate, mycelium white to buff, reverse red brown and spreading as soluble pigment. On MEA 25-38 mm in diameter, velutinous, low; mycelium white, inconspicuous; reverse dull brownish-yellow with yellow soluble pigment. On CDA 50-65 mm in diameter, thin, floccose, orange white with few pale greenish conidial heads; reverse orange. Heads radiate to columna. Stipes smooth, brown at maturity, $30-200 \times 4.1-5.3 \mu m$. Vesicles flask-shaped, hemispherical, 5.0-12.4 µm in diameter. Metulae covering the upper half of the vesicle, $4.1-6.3 \times 2.5-3.5 \ \mu m$. Phialides dehiscing rapidly, 4.5-8.0 \times 2.0-3.5 μm . Conidia globose, smooth to rugulose, sparce, dull green to grey green, 3.0-4.0 µm in diameter. Ascomata cleistothecia globose, brown red, hülle cells yellowish, ascospores red, smooth, with short crest, tow of them obvious, two quite indistinct, 4.5-5.5 \times 3.0-4.5 μ m.

The fungus has been isolated from the seeds of lentil, fenugreek and faba bean.

A. unguis (Emile Wiel and Gaudin) Thom and Raper (E. unguis Malloch and Cain) (Fig. 17).

On CYA colony diameter 20-37 mm, at 37°C 12-18 mm, velutinous, irregularly sulcate, with floccose central overlay of white mycelium, greyish-green becoming olive green to brown with age; exudate absent; cleistotheacia absent; reverse dull brown to red brown. On CY20S 23-32 mm in diameter, olive green, reverse

111

orange brown to black with age; other characters similar to those on CYA. On MEA 25-40 mm in diameter, sparce growth, plane, grey green to dark green; mycelium white, inconspicuous; exudate absent; reverse cream to pale yellow. On CDA 20-36 mm in diameter, other characters similar to those on CYA Heads radiate to loosely columnar on CYA, definitely columnar on MEA and with sterile, thick, white spicular hyphae arising above the conidial heads. Stipes simple or dichatomously branched, smooth, straight or sinuous, colorless to dull brown, $150\text{-}300 \times 3,3\text{-}6.0 \ \mu\text{m}$. Vesicles spathulate, hemispherical to flask-shaped, 5.0-12.0 µm wide. Metulae covering the upper half to the two third of the vesicle, $4.0\text{--}8.0 \times 2.5\text{--}3.5 \ \mu\text{m}$. Phialides ampulliform, $4.5-9.0 \times 2.0-3.5 \mu m$. Conidia Gglobose, smooth to echinulate or rugulose, 30-4.1 µm in diameter. Ascomata cleistothecia only on MEA, mature one not observed, 150-250 µm; ascospores lenticular, purple red, two equatorial crest, smooth, 4.0- $5.5 \times 3.0-3.5 \ \mu m.$

The fungus has been reported on the seeds of guar, lentil, fenugreek and faba bean.

A. violaceo-brunneus Samson and W. Gams (E. violacea (Fennell and Raper) Mallocch and Cain) (Fig. 18).

On CYA colony 45-60 mm at 37°C 33-40 mm; conidial structures not observed; exudate absent; reverse red to dark or brown red. On CY20S 45-47 mm in diameter, green and yellow in centre due to cleistothecia, reverse tan brown, other characters similar to those on MEA. On MEA 45-53 mm in diameter, grey green, yellowish in the centre due to abundant cleistothecia; wrinkled, umbonate; margin irregular, exudate present; reverse yellow brown. On CDA 62-70 mm in diameter, grey green to dark green, radially wrinkled; reverse reddish-brown in the centre to bluish-black or red towards the margin. Heads columnar. Stipes thick, sinuous, smooth, colorless to pale brown, 50-200 \times 4.1-5.3 $\mu m.$ Vesicles globose to hemispherical, 8.2-12.3 µm in diameter. Metulae bearing two phialides, $6.2-7.8 \times 2.5-3.3 \ \mu m$. Phialides ampulliform with short neck, 5.2-6.5 \times 2.5-3.1 μm . Conidia globose to elliptical, smooth to finely roughened, 3.1-5.0 µm. Ascomata cleistothecia globose, 120-280 μm ; hülle cells globose to subglobose dense, 12.2-22.4 μm in

diameter; ascospores violet blue, lenticular, echinulate, two narrow equatorial ridges, $5.0-6.2 \times 4.4-5.0 \mu m$.

The fungus has been has been isolated form the seeds of guar, lentil, lupine, pea, faba bean, and cowpea

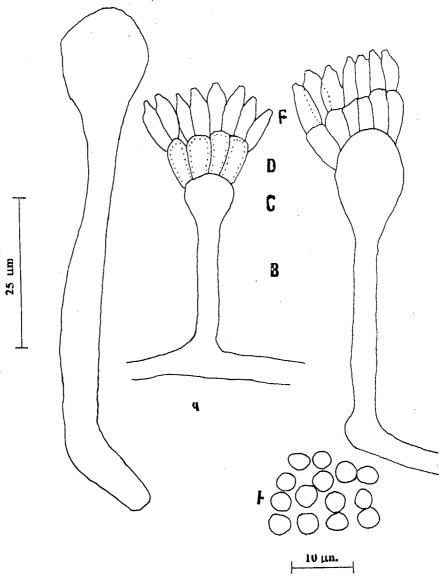


Fig. 1: Aspergillus caespitosus.

(A) Foot cells (B) Stipes (C) Vesicles.

(D)Metulae (E) Phialides (F) Conidia.

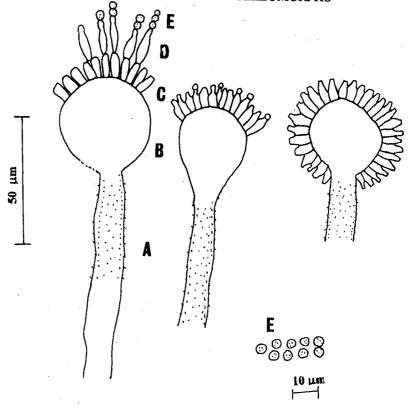


Fig. 2: Aspergillus flavus var. flavus.

- (A) Stipes (B) Vesicles (C) Metulae.(D) Phialides (E) Conidia.

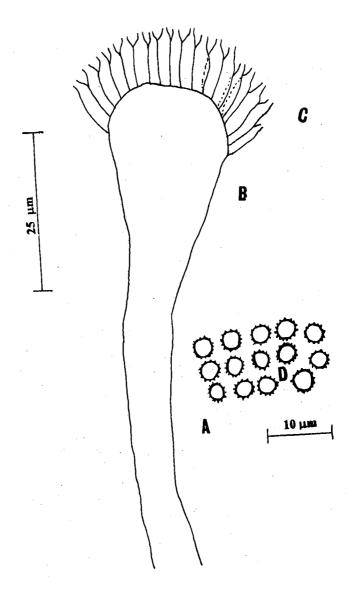


Fig. 3: Aspergillus fumigatus.

(A) Stipe (B) Vesicle.

(C) Phialides (D) Conidia.

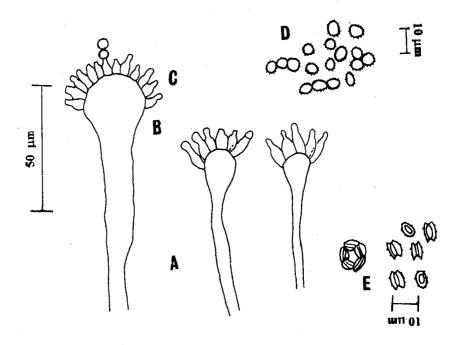


Fig. 4: Aspergillus hollandicus (Teleomorph: Eurotium amstelodami).

- (A) Stipes (B) Vesicles (C) Phialides.
- (D) Conidia (E) Ascus and ascospores.

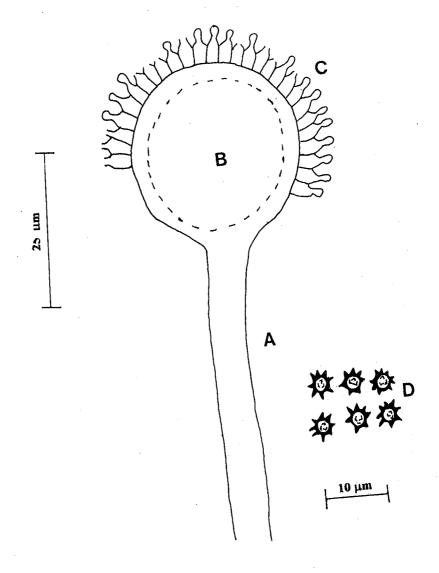


Fig. 5: Aspergillus japonicus var. aculeatus.
(A) Stipe (B) Vesicle.
(C) Phialides (D) Conidia.

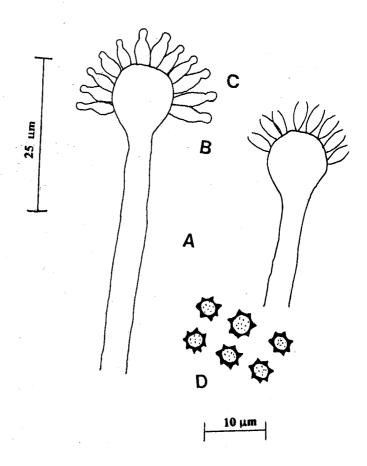


Fig. 6: Aspergillus japonicus var. japonicus.
(A) Stipes (B) Vesicles.
(C) Phialides (D) Conidia.

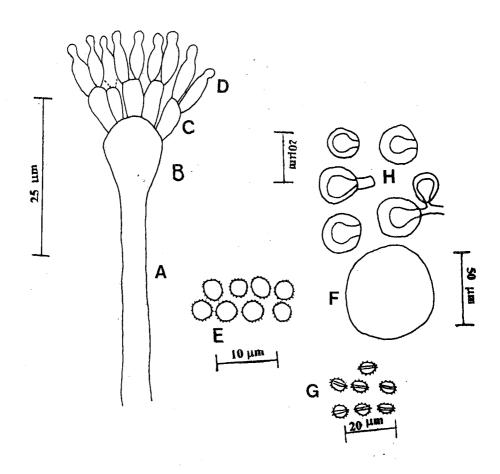


Fig. 7: Aspergillus nidulans var. echinulata (Teleomorph: Emericella nidulans var. echinulata).

- (A) Stipe (B) Vesicle (C) Metulae (D) Phialides (E) Conidia.
- (F) Cleistothecium (G) Ascospores (H) Hülle cells.

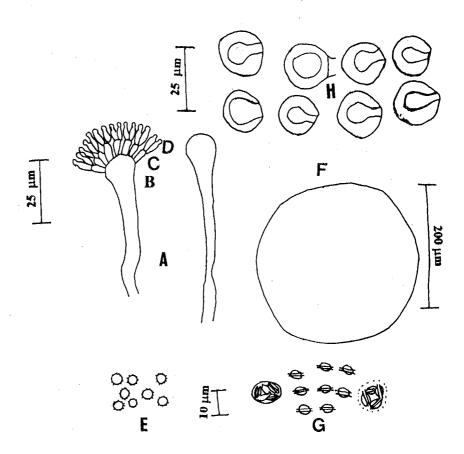


Fig. 8: Aspergillus nidullelus var. nidulans (Teleomorph: Emericella nidulans var. nidulans).

(A) Stipes (B) Vesicles (C) Metulae (D) Phialides (E) Conidia (F) Cleistothecium (G) Asci and ascospores (H) Hülle cells.

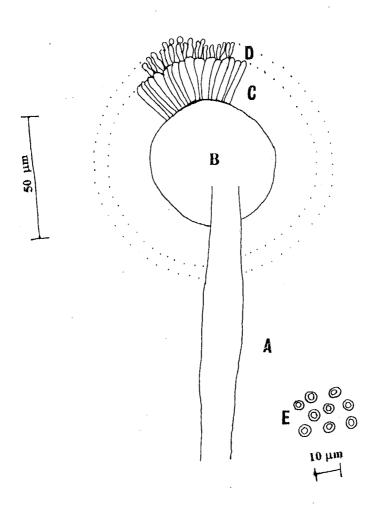


Fig. 9: Aspergillus niger.

(A) Stipe (B) Vesicle (C) Metulae.

(D) Phialides (E) Conidia.

Ò

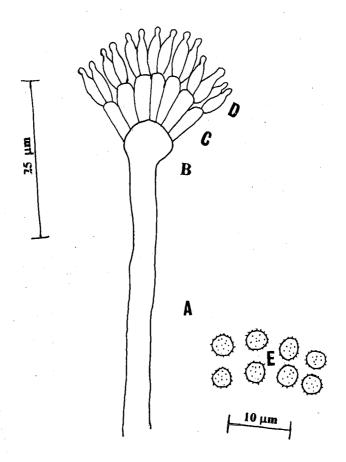


Fig. 10: Aspergillus niger var. awamori.

(A) Stipe (B) Vesiele (C) Metulae.

(D) Phialides (E) Conidia.

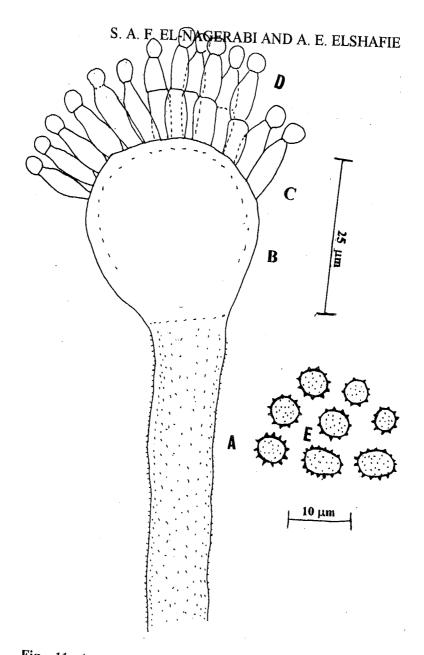


Fig. 11: Aspergillus oryzae.

(A) Stipe (B) Vesicle (C) Metulae.

(D) Phialides (E) Conidia.

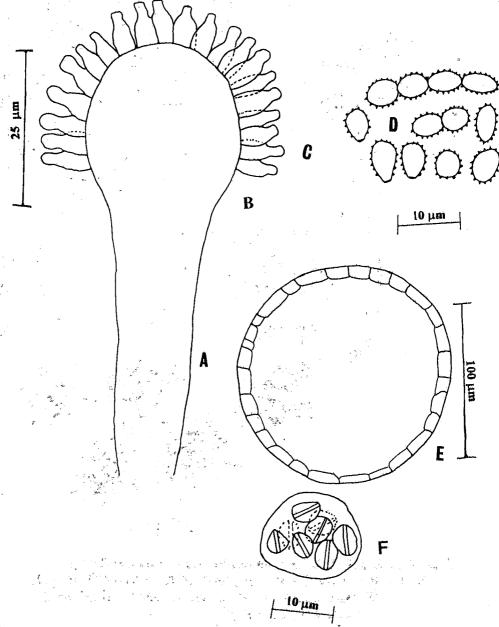


Fig. 12: Aspergillus rubrobrunneus (Teleomorph; Eurotium rubrum).

(A) Stine (B) Vesicle (C) Phialides (D) Conidia

(A) Stipe (B) Vesicle (C) Phialides (D) Conidia.
(E) Cleistothecium (F) Ascus and ascospores.

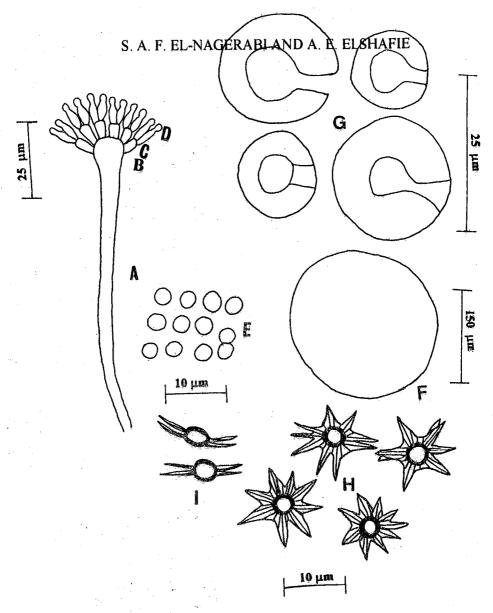


Fig. 13: Aspergillus stellifer (Teleomorph: Emericella variecolor).

(A) Stipe (B) Vesicle (C) Metulae (D) Phialides (E)Conidia

(F)Cleistothecium (G) Hülle cells (H)Ascospores(Face view)

(1) Ascospores (Side view).

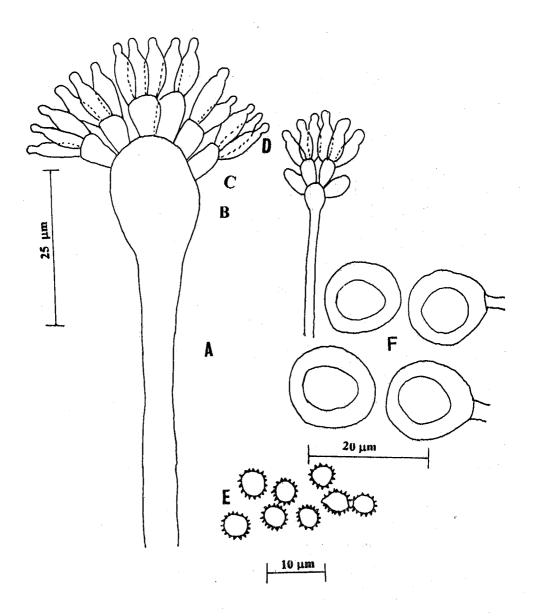


Fig. 14: Aspergillus sydowii.

71

- (A) Stipes (B) Vesicles (C) Metulae (D) Phialides
- (E) Conidia (F) Hülle cells.

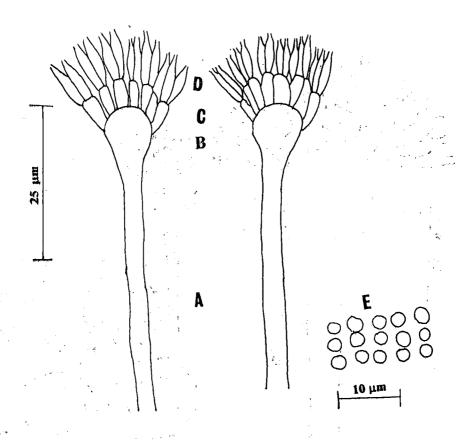


Fig. 15: Aspergillus terreus yar. terreus.

(A) Stipes (B) Vesicles (C) Metulae.

(D) Phialides (E) Conidia.

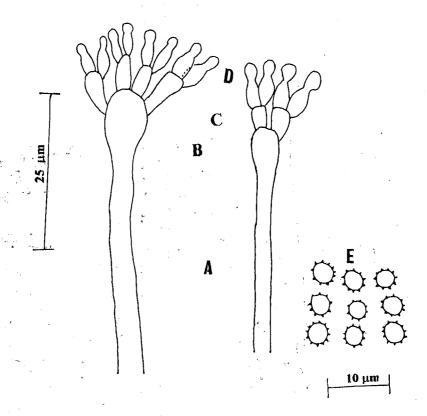


Fig. 16: Aspergillus tetrazonus.

- (A) Stipes (B) Vesicles (C) Metulae.
 (D) Phialides (E) Conidia.

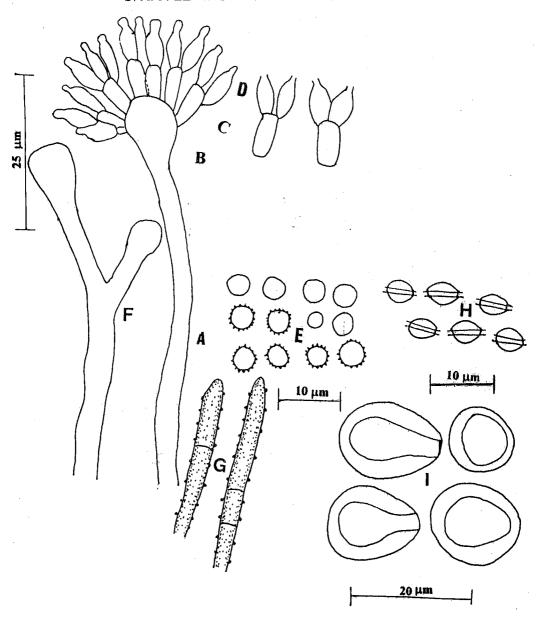


Fig. 17: Aspergillus unguis (Teleomorph: Emericella unguis).

(A) Stipe (B) Vesicle (C) Metulae (D) Phialides (E) Conidia.

(F) Dichotomous stipe (G) Sterile hyphae (H) Ascospores.

(I) Hülle cells.

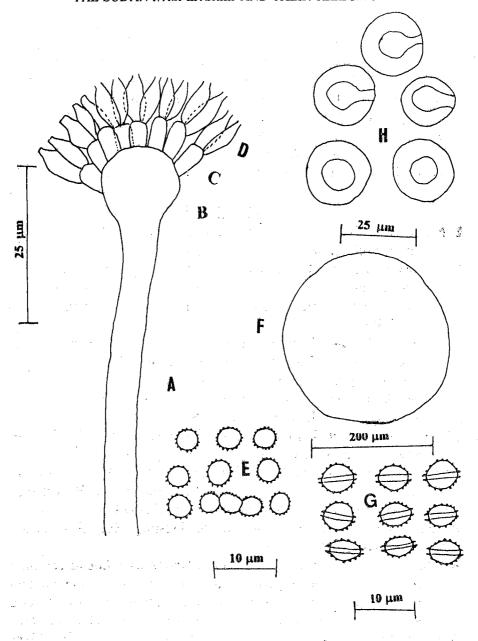


Fig. 18: Aspergillus violaceo- brunneus (Teleomorph: Emericella violacea).

(A) Stipe (B) Vesicle (C) Metulae (D) Phialides (E) Conidia. (F) Cleistothecium (G) Ascospores (H) Hülle cells.

ACKNOWLEDGEMENTS

)

1)

The authors wish to express their appreciation and gratitude to Prof. E. M. M. Abdel Bari of the University of Qatar for financial support and to Department of Botany and Computer Centre of University of Khartoum for providing space and facilities to carry this research. An acknowledgement should be directed to Dr. S. A. Ibrahim, Dr. S. Eltigani, Miss R. M. El-Hassan, Miss. M. J. Sharef El-Din and Miss I. Fadalalla for their advice and encouragement.

REFERENCES

- Blakeslee, A. F., 1915. Linder's roll tube method for separating cultures. Phytopathology, V. 5, p. 68-69.
- Christensen, C. M., 1963. Influence of small difference in moisture content upon the invasion of hard red winter wheat by Aspergillus restrictus and A. repens. Cereal Chem., V.40, p.385-390.
- El-Kady, I. A., and Youssef, M. S., 1993. Survey of mycoflora and mycotoxins in Egyptian soybean seeds. Journal of Basic Microbiology, V.33(6), p. 371-378.
- Elshafie, A. E., 1985. Taxonomic studies of seed-borne fungi of the Sudan, 1: Drechslera. Sudan J. of Sci., V.I, p. 62-84.
- Elshafie, A. E., 1986. Taxonomic studies on seed-borne fungi of the Sudan II: Curvularia. Sudan J. Sci., V.II, p. 51-70.
- Hussain, S. S., Hassan, S., and Khan, B. A., 1989. Seed-borne mycoflora of soybean in the Northern West Frontier Province of Pakistan. Sarhad J. of Agric., V.5(4), p. 421-424.
- ISTA., 1966. International rules for seed testing. Seed Science and Technology, V.4, p. 3-49.
- Klich, M. A., and Pitt, J. I., 1988. A laboratory guide to common Aspergillus species and their teleomorphs. Commnwealth Scientific and Institute Research Organization, Division of Food Processing, North Ryde, New South Wales, Australia. 116 pp.
 - Moslem, M. A., and Parvez, S., 1993. Seed-borne fungi of lens esculentus, Hordeum vulgare and Triticum aestivum from Saudi Arabia. International Journal of Tropical Plant Diseases, V.11(1), p. 99-105.

- Moubasher, A. H., 1993. Soil Fungi in Qatar and Other Arab countries. The Centre for Scientific and Applied Research, University of Qatar, Doha Modern Printing Press, 1st ed., 566pp.
- Pitt, J. I., 1973. An appraisal of identification methods for Penicillium species: Novel taxonomic criteria based on temperature and water relations. Mycologia, V.65, p. 1135-1157
- Pitt, J. I., and Hocking, A. D., 1985. Fungi and food spoilage. Sydney, Australia, Academic Press.
- Raper, K. B., and Fennell, D. I., 1965. The genus Aspergillus. The William and Wilkins Company, Baltimore, USA. 686 p.
- Singh, K., Frisval, J. C., Thrane, U. and Mathur, S. M., 1991. An illustrated manual on identification of some seed-borne Aspergilli, Fusaria, Penicillia and Their mycotoxins. Danish Government Institute of Seed Pathology for Developing Countries, 1st ed. 133 p.
- Tarr, S. A. J., 1955. The Fungi and Plant Diseases of the Sudan. Commonwealth Mycological institute, Kew, Surrey, England, 127 p.
- Tarr, S. A. J., 1963. A supplementary list of Sudan fungi and plant diseases. Mycological paper No. 85, Commonwealth Mycological Institute, Kew, Surrey, England.
- Zohri, A. A., Abdel Gawad, K., 1992. Studies on mycoflora and mycotoxins of cowpea cultivars. Korean J. Mycol., V.20(3), p.252-258.